

# Product Sheet

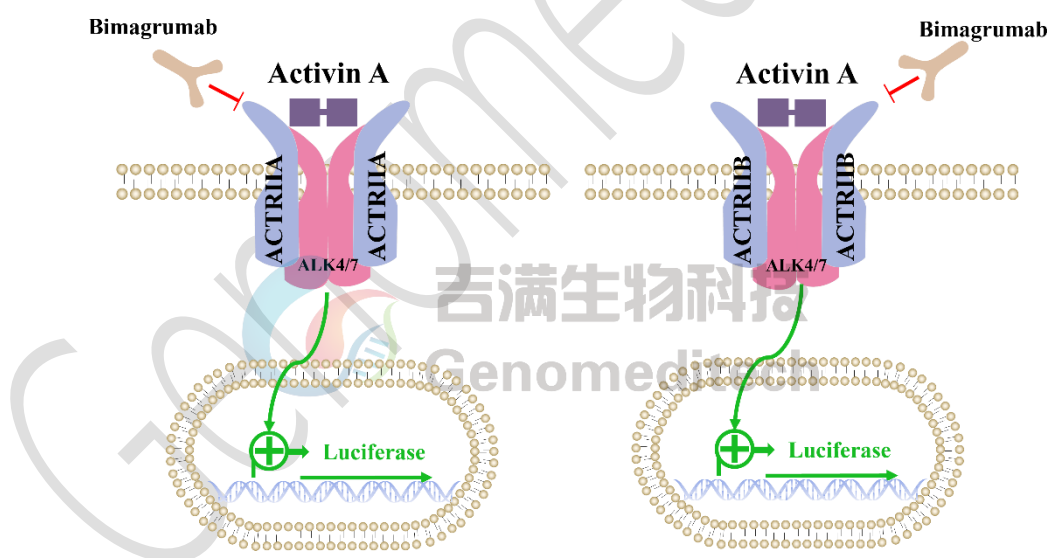
## Activin A Reporter Cell Line

Catalog number: GM-C26082

Version 3.3.1.241128

The Activin type II receptors (ActRII) include two distinct receptors, ActRIIA (ACVR2A) and ActRIIB (ACVR2B), which are members of the transforming growth factor-beta (TGF- $\beta$ ) receptor family. They serve as type II receptors for other TGF- $\beta$  family members, including Activin A, Activin B, BMP7, BMP9, BMP10, GDF1, GDF8 (myostatin), GDF11, and Nodal. ALK4 and ALK7 are the primary type I TGF- $\beta$  receptor family members for Activin A and Activin B, respectively. Bimagrumab is a human monoclonal antibody that can block Activin type II receptors (ActRII), thereby regulating skeletal muscle growth. Research has shown that Bimagrumab has an affinity for both ActRIIB and ActRIIA, but its affinity for ActRIIB is at least 200 times stronger than for ActRIIA.

Activin A Reporter Cell Line is a clonal stable cell line with signal-dependent expression of a luciferase reporter gene constructed using lentiviral technology, and it endogenously expresses ACTRII gene. When Activin A or GDF-8 binds to ACTRII, it activates downstream signaling pathways, leading to the expression of luciferase. Blockade antibodies can inhibit this signal transmission. The luciferase activity measurement indicates the activation level of the signaling pathway and can thus be used to evaluate the in vitro effects of drugs related to this signaling pathway.



## Specifications

<b>Quantity</b>	5E6 Cells per vial,1 mL
<b>Product Format</b>	1 vial of frozen cells
<b>Shipping</b>	Shipped on dry ice
<b>Storage Conditions</b>	Liquid nitrogen immediately upon receipt

<b>Recovery Medium</b>	DMEM+10% FBS+1% P.S
<b>Growth medium</b>	DMEM+10% FBS+1% P.S+4 µg/mL Blasticidin
<b>Note</b>	None
<b>Freezing Medium</b>	90% FBS+10% DMSO
<b>Growth properties</b>	Adherent
<b>Growth Conditions</b>	37°C, 5% CO <sub>2</sub>

<b>Mycoplasma Testing</b>	The cell line has been screened to confirm the absence of Mycoplasma species.
<b>Safety considerations</b>	Biosafety Level 2
<b>Note</b>	It is recommended to expand the cell culture and store a minimum of 10 vials at an early passage for potential future use.

## Materials

<b>Reagent</b>	<b>Manufacturer/Catalogue No.</b>
DMEM	Gibco/C11995500BT
Fetal Bovine Serum	Cegrogen biotech/A0500-3010
Pen/Strep	Thermo/15140-122
Blasticidin	Genomeditech/ <a href="#">GM-040404</a>
Human Activin A Protein; His Tag	Genomeditech/ <a href="#">GM-87616RP</a>
Anti-ACVR2B hIgG1 Antibody(Bimagrumab)	Genomeditech/ <a href="#">GM-51148AB</a>
GMOne-Step Luciferase Reporter Gene Assay Kit	Genomeditech/ <a href="#">GM-040503</a>

## Figures

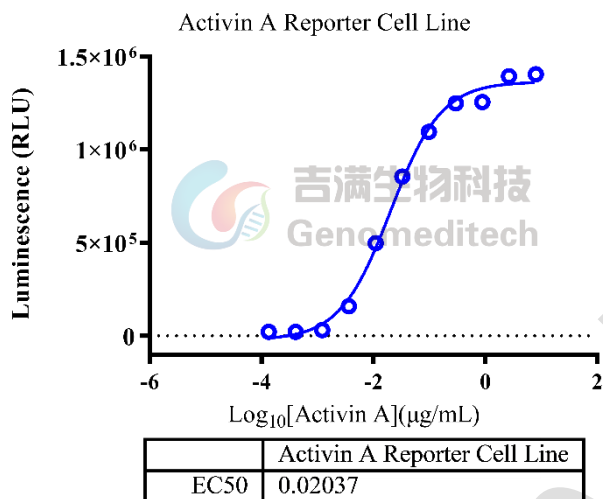


Figure 1 | Response to Activin A Protein. Activin A Reporter Cell Line (Cat. GM-C26082) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Human Activin A Protein (Cat. GM-87616RP) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The maximum induction fold was approximately [63.2]. Data are shown by drug mass concentration.

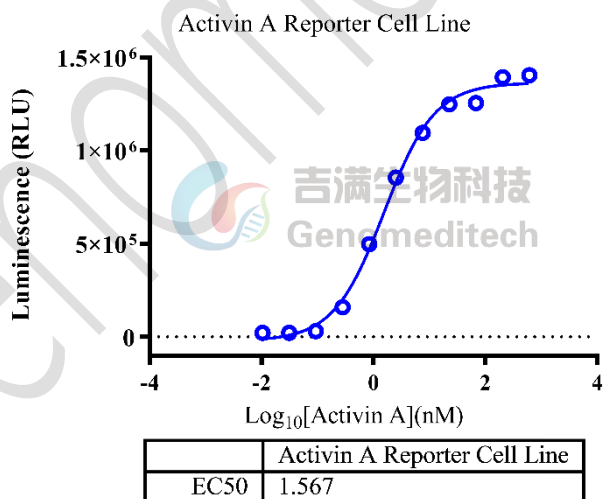


Figure 2 | Response to Activin A Protein. Activin A Reporter Cell Line (Cat. GM-C26082) at a concentration of 1.5E4 cells/well (96-well format) was stimulated with serial dilutions of Human Activin A Protein (Cat. GM-87616RP) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. GM-040503). The maximum induction fold was approximately [63.2]. Data are shown by drug molar concentration.

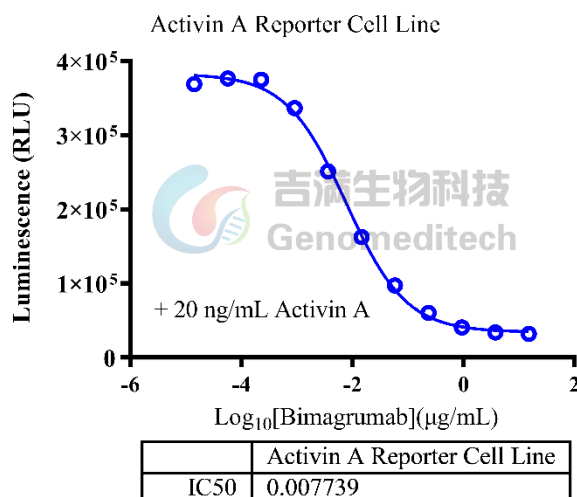


Figure 3 | Response to Anti-ACVR2B hIgG1 Antibody(Bimagrumb). Serial dilutions of the Anti-ACVR2B hIgG1 Antibody(Bimagrumb) (Cat. [GM-51148AB](#)) was incubated with 1.5E4 cells/well of the Activin A Reporter Cell Line (Cat. GM-C26082) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). Subsequently, the Human Activin A Protein (Cat. [GM-87616RP](#)) at a concentration of 2 ng/well was added, and the coculture proceeded for an additional 6 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). The results indicated maximum blocking folds of approximately [11.6]. Data are shown by drug mass concentration.

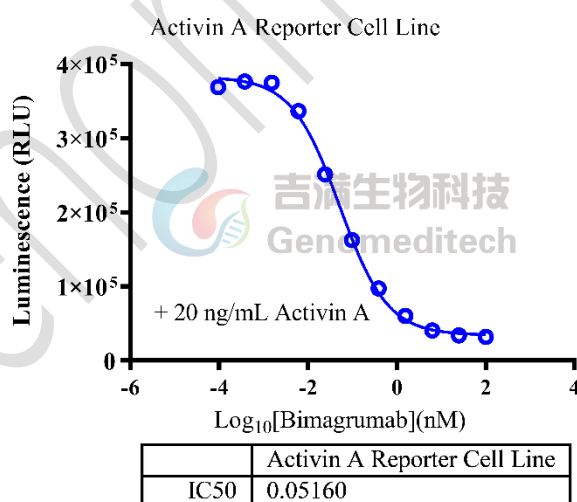


Figure 4 | Response to Anti-ACVR2B hIgG1 Antibody(Bimagrumb). Serial dilutions of the Anti-ACVR2B hIgG1 Antibody(Bimagrumb) (Cat. [GM-51148AB](#)) was incubated with 1.5E4 cells/well of the Activin A Reporter Cell Line (Cat. GM-C26082) in a 96-well plate for 1 hour in assay buffer (DMEM + 1% FBS + 1% P.S). Subsequently, the Human Activin A Protein (Cat. [GM-87616RP](#)) at a concentration of 2 ng/well was added, and the coculture proceeded for an additional 6 hours. Firefly luciferase activity is then measured using the GMOne-Step Luciferase Reporter Gene

Assay Kit (Cat. [GM-040503](#)). The results indicated maximum blocking folds of approximately [11.6]. Data are shown by drug molar concentration.

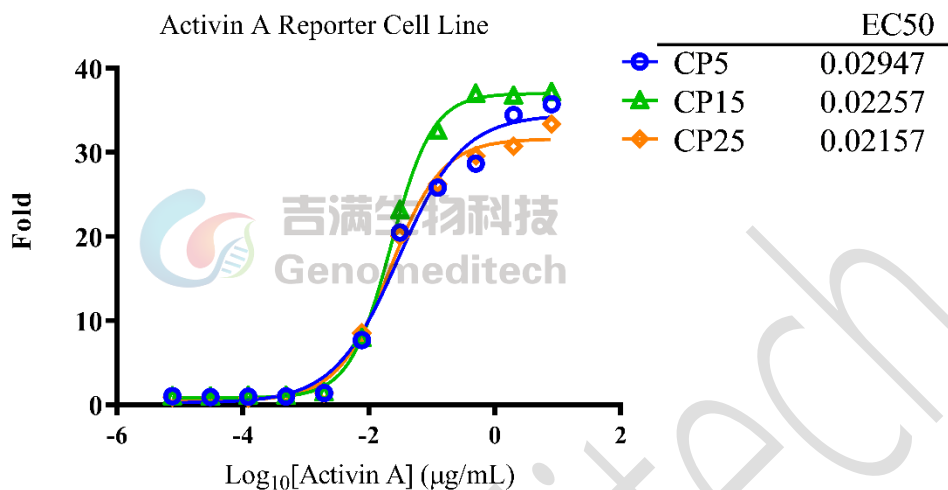


Figure 5 | The passage stability of response to Human Activin A Protein. The passage 5, 15 and 25 of Activin A Reporter Cell Line (Cat. [GM-C26082](#)) at a concentration of 1.5E4 cells/well (96-well format) were stimulated with serial dilutions of Human Activin A Protein (Cat. [GM-87616RP](#)) in assay buffer (DMEM + 1% FBS + 1% P.S) for 7 hours. The firefly luciferase activity was measured using the GMOne-Step Luciferase Reporter Gene Assay Kit (Cat. [GM-040503](#)). Data are shown by drug mass concentration.

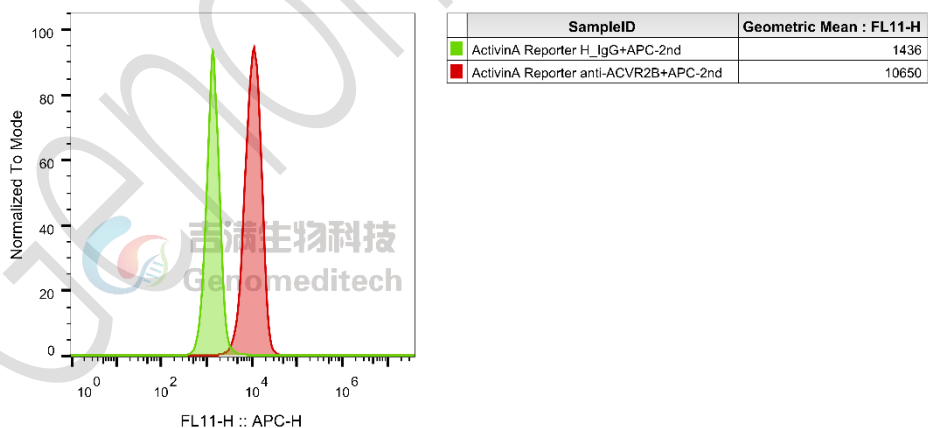


Figure 6 | Activin A Reporter Cell Line (Cat. [GM-C26082](#)) was determined by flow cytometry using Anti-ACVR2B hIgG1 Antibody(Bimagramab) (Cat. [GM-51148AB](#)).

## Cell Recovery

Recovery Medium: DMEM+10% FBS+1% P.S

To insure the highest level of viability, thaw the vial and initiate the culture as soon as possible upon receipt. If upon arrival, continued storage of the frozen culture is necessary, it should be stored in liquid nitrogen vapor phase and not at  $-70^{\circ}\text{C}$ . Storage at  $-70^{\circ}\text{C}$  will result in loss of viability.

- Thaw the vial by gentle agitation in a  $37^{\circ}\text{C}$  water bath. To reduce the possibility of contamination, keep the O-ring and cap out of the water. Thawing should be rapid (approximately 2 - 3 minutes).
- Remove the vial from the water bath as soon as the contents are thawed, and decontaminate by dipping in or spraying with 70% ethanol. All of the operations from this point on should be carried out under strict aseptic conditions.
- Transfer the vial contents to a centrifuge tube containing 5.0 mL complete culture medium and spin at approximately  $176 \times g$  for 5 minutes. Discard supernatant.
- Resuspend cell pellet with the recommended recovery medium. And dispense into appropriate culture dishes.
- Incubate the culture at  $37^{\circ}\text{C}$  in a suitable incubator. A 5%  $\text{CO}_2$  in air atmosphere is recommended if using the medium described on this product sheet.

## Cell Freezing

Freezing Medium: 90% FBS+10% DMSO

- Centrifuge at  $176 \times g$  for 3 minutes to collect cells.
- Resuspend the cells in pre-cooled freezing medium and adjust the cell density to  $5 \times 10^6$  cells/mL.
- Aliquot 1 mL into each vial.
- Place the vial in a controlled-rate freezing container and store at  $-80^{\circ}\text{C}$  for at least 1 day, then transfer to liquid nitrogen as soon as possible.

## Cell passage

Growth medium: DMEM+10% FBS+1% P.S+4  $\mu\text{g/mL}$  Blasticidin

For the first 1 to 2 passages post-resuscitation, use the recovery medium. Once the cells have stabilized, switch to a growth medium.

- Subculturing is necessary when the cell density reaches 80%. It is recommended to perform subculturing at a ratio of 1:3 to 1:4 every 2-3 days. Ensure that the density does not exceed 80%, as overcrowding can lead to reduced viability due to compression.
- Remove and discard culture medium.
- Briefly rinse the cell layer with PBS to remove all traces of serum that contains trypsin inhibitor.
- Add 1.0 mL of 0.25% (w/v) Trypsin-EDTA solution to dish and observe cells under an inverted microscope until cell layer is dispersed (usually within 30 to 60 seconds at  $37^{\circ}\text{C}$ ).
- Note: To avoid clumping do not agitate the cells by hitting or shaking the flask while waiting for the cells to detach. Cells that are difficult to detach may be placed at  $37^{\circ}\text{C}$  to facilitate dispersal.
- Add 2.0 mL of growth medium to mix well and aspirate cells by gently pipetting.

- g) After centrifugation, resuspend the pellet and add appropriate aliquots of the cell suspension to new culture vessels.
- h) Incubate cultures at 37°C.

**Subcultivation Ratio: A subcultivation ratio of 1:3 - 1:4 is recommended**

**Medium Renewal: Every 2 to 3 days**

## Notes

- a) Upon initial thawing, a higher number of dead cells is observed, which is a normal phenomenon. Significant improvement is seen after adaptation. Once the cells reach a stable state, the number of dead cells decreases after subculturing and the cell growth rate becomes stable.
- b) Ensure that the cell density does not exceed 80%, as overcrowding may lead to reduced viability due to compression.

## Related Products

GCGR	
<a href="#">H_GCGR Reporter CHO-K1 Cell Line</a>	<a href="#">H_GCGR Reporter HEK-293 Cell Line</a>
<a href="#">H_GCGR CHO-K1 Cell Line</a>	<a href="#">H_GCGR HEK-293 Cell Line</a>
<a href="#">Mouse_GCGR HEK-293 Cell Line</a>	
<a href="#">Anti-H_GCGR hIgG2 Antibody(volagidemab)</a>	
GLP1R	
<a href="#">H_GLP1R Reporter CHO-K1 Cell Line</a>	<a href="#">H_GLP1R Reporter HEK-293 Cell Line</a>
<a href="#">Cynomolgus_GLP1R HEK-293 Cell Line</a>	<a href="#">H_GLP1R CHO-K1 Cell Line</a>
<a href="#">H_GLP1R HEK-293 Cell Line</a>	<a href="#">Mouse_GLP1R HEK-293 Cell Line</a>
<a href="#">Anti-GLP1R hIgG1 Antibody(mAb-36986)</a>	<a href="#">Anti-H_GLP1R hIgG1 Antibody(glutazumab)</a>
FGF21:FGFR	
<a href="#">H_FGF21 Reporter HEK-293 Cell Line</a>	
CALCA(CGRP): CALCRL RAMP	
<a href="#">H_CALCRL RAMP1 Reporter HEK-293 Cell Line</a>	<a href="#">Cynomolgus_CALCRL RAMP1 HEK-293 Cell Line</a>
<a href="#">H_CALCRL RAMP1 CHO-K1 Cell Line</a>	<a href="#">H_CALCRL RAMP1 HEK-293 Cell Line</a>
<a href="#">H_CALCRL RAMP2(AM1) CHO-K1 Cell Line</a>	<a href="#">H_CALCRL RAMP3(AM2) CHO-K1 Cell Line</a>
<a href="#">Anti-CALCRL RAMP1 hIgG2 Antibody(Erenumab)</a>	
GIP:GIPR	
<a href="#">H_GIPR Reporter CHO-K1 Cell Line</a>	<a href="#">H_GIPR Reporter HEK-293 Cell Line</a>
<a href="#">H_GIPR Reporter HEK-293 DDX35TM Cell Line</a>	<a href="#">Cynomolgus_GIPR HEK-293 Cell Line</a>
<a href="#">H_GIPR CHO-K1 Cell Line</a>	<a href="#">H_GIPR HEK-293 Cell Line</a>
<a href="#">Mouse_GIPR HEK-293 Cell Line</a>	
<a href="#">Anti-H_GIPR hIgG1 Antibody(AMG-133)</a>	
ACVR2A: ACTRIIB: Active A	
<a href="#">ACVR2A KO HEK-293 Cell Line</a>	<a href="#">H_ACVR2A Reporter Cell Line</a>
<a href="#">H_ACVR2B Reporter Cell Line</a>	<a href="#">ACVR2B KO HEK-293 Cell Line</a>

H_ACVR2A HEK-293(ACVR2B KO) Cell Line	H_ACVR2B HEK-293(ACVR2A KO) Cell Line
Anti-ACVR2B hIgG1 Antibody(Bimagrumab)	Anti-ACVR2B hIgG1 Antibody(Fab-17G05)
Anti-ACVR2B mIgG2a Antibody(Bimagrumab)	Anti-H_ACVR2B hIgG1 Reference Antibody(Bimbio)
Biotinylated Human ACVR2B Protein; His-Avi Tag	Biotinylated Mouse ACVR2A Protein; His-Avi Tag
Biotinylated Mouse ACVR2B Protein; His-Avi Tag	Human Activin A Protein; His Tag
Human Activin B Protein; His Tag	Human ACVR2A Protein; hFc Tag
Human ACVR2A Protein; His Tag	Human ACVR2B Protein; hFc Tag
Human ACVR2B Protein; His Tag	Mouse ACVR2B Protein; His Tag
<b>AMY: CALCR RAMP</b>	
H_CALCR RAMP3(AMY3) Reporter CHO-K1 Cell Line	H_CALCR Reporter CHO-K1 Cell Line

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